# Method Development for the Analysis of Residual Solvents and Terpenes in one run using headspace GC-MS/FID

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Aailent Trusted Answers

# Introduction

There is a growing need for the analysis of residual solvents and terpenes in cannabis and cannabinoid concentrates. These methods must assure safety and quality and the list of terpenes and residual solvents that are to be analyzed is continuously being increased by state and local governing bodies. A common configuration for these analyses connects headspace sampling techniques with GC or GC-MS systems. Agilent is collaborating with major laboratories to rapidly develop high-end instrument configurations and methodologies that are robust, accurate and precise while still allowing for easy to implement workflows in high productivity laboratories. The Agilent headspace-GC-MS systems for the analysis of residual solvents and terpenes includes the 7697A headspace auto-sampler, the 7890B GC and the 5977B mass spectrometer operated on MassHunter software. This presentation will discuss the Agilent systems and methodologies for the analysis of terpenes and residual solvents in cannabis and cannabinoid samples.

# **Experimental**

#### Workflow for samples and standards:

10 µL standard is placed in a 20mL headspace vial and capped. A 5 or 10 point calibration curve is created from the standards.

As plant and extract material does not dissolve in solvent, a full evaporation headspace technique (FET) is used for quantitation.

A part of the flower weighing 1.0 gram is frozen, followed by grinding to ensure a representative sample. 10-30 mg of the flower is then weighed into a headspace vial and capped.

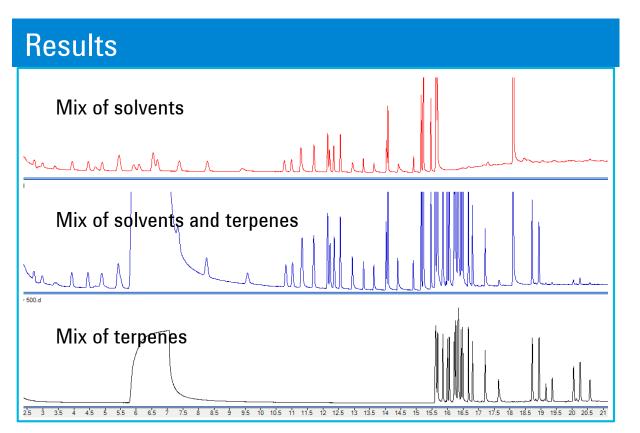
For different species of samples:

plant, extracts, etc. Use 5-7 mg +/- 15 mg place into 20 mL headspace vial, evaluate against curve.

Run QC to verify performance

## **Experimental**

Analytical Column Agilent 122-1334UI DB- 624 Ultra Inert 20 °C—260 °C (360 °C n Dut Initial) Pressure How Average Velocity Holdup Time	c): 30 m x 250 µm x 1.4 µm Front SS Inlet He Aux EPC 1 35 °C 11.922 psi 1.0 mL/min 24.63 cm/sec 0.97842 min	Column In Out (Initial) Pressure Flow Average Velocity FID Restrictor -60 °C—350 °C (350 °C Column In Out	<ul> <li>c): 1.7 m x 150 μm x 0 μm Deactivated fused silica Aux EPC 1 He MSD 35 °C 0.47017 psi 1.15 mL/min 162.94 cm/sec</li> <li>c): 0.7 m x 250 μm x 0 μm Deactivated fused silica Aux EPC 1 He Front Detector FID</li> </ul>
		(Initial)	35 °C



Pressure

Average Velocity

Flow

0.18046 psi

0.5 mL/min

17.439 cm/sec

100.

103.

91.

104.5

100.8

100.9

85. 115.2

Figure 1. MS Chromatograms residual solvent mix only (top), a mix of both (middle) and terpene only mix(bottom), 29.0 minute cycle times

						_
ation Curve → \$ " ■ ■ Type: Quadratic → Origin: L → Weight: 1/x → ISTD QC		Toluene Results				
ene - 8 Levels Used, 8 Points, 8 Points Used, 0 QCs 105 [ y = 425651 * x <sup>2</sup> - 7114 63874 * x - 127 224187	RT	Resp.	М	Calc. Conc.	Final Conc.	Aco
1 R^2 = 0.99921038 .75−1 Type:Quadratic, Origin:Include, Weight:1/x	15.053	3 0				
Calibration, 8 levels. 0.5 $\mu$ g/g to 500.0 $\mu$ g/g	14.08	47912		500.3649	500.3649	
Toluono	14.082	24864		291.1016	291.1016	
Toluene	14.082	2 720203		95.2456	95.2456	
	14.082	392572		53.4954	53.4954	
<sup>25</sup> <b>-</b> <sup>2</sup> 0 000						

# Compound list, target ion, and retention times

NAME	m/z	RT	NAME	m/z	RT
propane	44	2.221	Eucalyptol	81	16.435
Isobutane	43	2.72	.gammaTerpinene	93	16.486
Butane	43	2.988	terpinolene	136	16.672
Acetone	43	3.122	Linalool	69	16.803
Methanol	31	3.37	Isopulegol	81	17.206
Butane, 2-methyl-	57	3.94	Geraniol		17.637
Pentane	43	4.46		93	
Ethanol	31	4.67	Caryophyllene	133	18.722
Ethyl ether	74	4.9	Humulene	93	18.941
Butane, 2,2-dimethyl-	71	5.437	Nerolidol	93	19.159
Isopropyl Alcohol	45	5.88	Nerolidol 2	93	19.364
Acetonitrile	41	6.08	Guaiol	161	20.049
Methylene chloride	84	6.53	Caryophyllene oxide	93	20.259
Pentane, 2-methyl-	71	6.673	.alphaBisabolol	109	20.576
Pentane, 3-methyl-	57	7.38			
n-Hexane	57	8.27			
1-Propanol	31	9.35			
2-Butanone	72	10.75			
Ethyl Acetate	43	10.98			
Tetrahydrofuran	72	11.27			
Cyclohexane	84	11.7			
Benzene	78	12.14			
Ethane, 1,2-dimethoxy-	45	12.2			
Isopropyl acetate	43	12.34			
Heptane	71	12.55			
1-Butanol	56.1	12.929			
1,4-Dioxane	88	13.293			
Ethanol, 2-ethoxy-	59	13.62			
Pyridine	79	14.03			
Toluene	91.1	14.082			
1-Pentanol	55	14.4			
Formamide, N,N-dimethyl-	73	14.9			
Ethylbenzene	91	15.15			
m-p-Xylene	91.1	15.218			
o-xylene	106	15.46			
Dimethyl Sulfoxide	78	15.62			
N,N-Dimethylacetamide	87	15.66			
Cumene	120	15.67			
tetramethylene sulfone	120	15.675			
.alphaPinene	93	15.683			
Camphene	93	15.851			
.betaMyrcene	69	15.993			
.betaPinene	93	16.055			
delta -3 Caren3	93	16.218			
alpha terpinene	121	16.276			
Limonene	68	16.331			
p-Cymene	119	16.347			
.betaOcimene	79	16.356			
L					

7697A Headspace parameters		Fill Pressure (psi):	15
Carrier Control:	GC	Loop Fill Mode:	Custom
Instrument Conditions		Loop Ramp Rate (psi/min):	40
Oven Temperature (°C):	85	Loop Final Pressure (psi):	10
Loop Temperature (°C):	85	Loop Equilibration Time:	0.05
Transfer Line Temperature (°C):	100	Carrier Control Mode:	GC controls
Vial Equilibration (min):	20.00	Extraction	
Injection Duration (min):	0.50	Vent After Extraction:	ON
GC Cycle Time (min):	29.00	Post Injection Purge:	100 mL/min for
Vial Size:	20	3 min	
Vial Shaking:	Level 2	Acceptable Leak Check:	Default,
		0.2mL/m	
Front SS Inlet He		Thermal Aux 2 (MSD Transfer	Line)

Set point

#### Mode Split 140 °C Heater 19.92 psi Pressure 21 mL/min Total Flow Septum Purge Flow off Split Ratio 20 :1

7890B GC Column Summary	
Oven Temperature Program	

••••••••••••••••••••••••••••••••••••••				
(Initial)				
Hold Time				
#1 Rate				
#1 Value				
#1 Hold Time				
#2 Rate				
#2 Value				
#2 Hold Time				
Equilibration Time				

#### Front Detector FID Conditions

Makeup	He
Heater	300 °C
H2 Flow	40
mL/min	
Air Flow	400
mL/min	
Makeup Flow	25
mL/min	

	5977 MS Condition	-		
	Acquisition Mode	: Scan		
35 °C	Solvent Delay (minutes) :0.1			
10.0 min	Tune file:			
30 °C/min	D:\MassHunter\GCMS\1\5977\atune.u			
180 °C	EM Setting mode Gain : 1.000000			
0 min	Ū			
40 °C/min	Normal or Fast Scanning : Normal Scanning			
260 °C				
5.0 min	[Scan Parameters]			
1 min	Start Time	0.1		
	Low Mass	30		
	High Mass	400		
He	Threshold	150		
300 °C	A/D Samples:	4		
40				
	MS Source Temp	300 C		

MS Quad Temp

150 C

280 °C

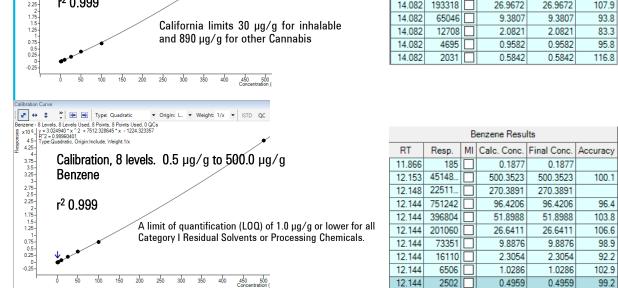


Figure 2. Example Residual Solvent calibration with California action limits for Benzene and Toluene 0.5  $\mu$ g/g to 500  $\mu$ g/g

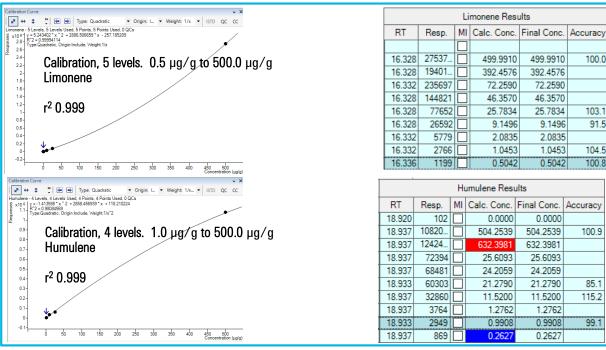


Figure 3. Example Terpene Calibration curves 0.5  $\mu$ g/g to 500 µg∕g

#### Conclusions

- Newest methodology enhancements allowing full scan acquisition of residual solvents and terpenes with a 29 minute run time and great separation using both FID and MS for quantitation.
- NIST spectral library search properly identifies all compounds
- At least two samples per hour can be run negating the need for two independent tests.
- Increased lab productivity

# References

- USP 467 Analysis of Residual Solvents. (2017) Agilent Technologies Application Note 5991-8032.
- Firor, R. L. Analysis of USP <467> Residual Solvents with Improved Repeatability Using the Agilent 7697A Headspace Sampler. (2012) Agilent Technologies Application Note 5990-7625.
- Honnold, R., Kubas, R., Macherone, A. (2017) Analysis of Terpenes in Cannabis Using the Agilent 7697A/7890B/5977B Headspace GC-MSD System: Faster Analysis Time = Greater Productivity. Agilent Technologies Application Note 5991-8499.

Agilent products and solutions are intended to be used for cannabis quality control and safety testing in laboratories where such use is permitted under state/country law.