CASE STUDY

# Overcoming the challenges of nitrosamine impurities in drugs

What pharmaceutical QA/QC laboratories need to know: Advanced GC-MS capabilities for cGMP nitrosamine testing

#### Why we need to analyze nitrosamine impurities in drugs

Unacceptable levels of nitrosamine impurities were first reported in June 2018 when Valsartan, an angiotensin Il receptor blocker containing a tetrazole group, was recalled due to the presence of N-nitrosodimethylamine (NDMA) contamination. Further nitrosamine impurities were subsequently detected in other medications belonging to a group of sartans, including: N-nitrosodiethylamine (NDEA), N-nitrosodiisopropylamine (NDIPA), N-nitrosoethylisopropylamine (NEIPA) and N-nitroso-N-methyl-4-aminobutyric acid (NMBA).<sup>1</sup> Nitrosamines are considered a matter of concern as the ICH M7 (R1)<sup>2</sup> Guideline classifies them as Class 1 impurities or mutagenic carcinogens and they are categorized as probable carcinogens by the International Agency for Cancer Research (IARC).<sup>3</sup> There are many possible sources of impurities in pharmaceutical substances; nitrites or secondary or tertiary amines can be present as unintentional contaminants of raw materials, reagents and solvents used during the production processes and they can result in the formation of nitrosamine impurities by reaction with a nitrosating agent (e.g., sodium nitrite (NaNO<sub>2</sub>).<sup>1</sup> Regulatory agencies all over the world have allowed a period of two years for manufacturers to review and make changes to their manufacturing processes to minimize nitrosamine impurities to the extent practically possible. During this transition period, interim limits are being applied to products based upon maximum daily intake.<sup>4</sup> Manufacturers of sartans with a tetrazole ring



have to implement a control strategy for N-nitrosamines and from April 2021, the batches of active pharmaceutical ingredient or drug substance they produce must not contain quantifiable levels (corresponding to less than 0.03 ppm) of the two principal N-nitrosamine impurities: NDMA and NDEA.

# Challenges of NDMA and other nitrosamine analysis in pharmaceuticals

The challenges of nitrosamine analysis in pharmaceuticals relate to sensitivity, selectivity, and compliance all in the light of obtaining reliable and timely results; high sensitivity must be achieved to fulfill the regulation requirements, and selectivity is also critical to avoid false positive noncompliant results.



"A challenge with headspace GC-based testing is carryover of residual organic solvents. We found that the TriPlus 500 headspace sampler fully addresses the carryover problem due to directly connecting to the column. TriPlus RSH autosampler is also a great option as it allows a quick switching between headspace and liquid injection modes, significantly increasing our throughput"

- Dr. Dujuan Lu (SGS, USA)

The U.S. Food and Drug Administration (FDA) has published several analytical methods that may be considered when determining nitrosamine content in active pharmaceutical ingredient (API) or finished pharmaceutical product (FPP).<sup>5</sup> These methods include both liquid chromatography (LC) and gas chromatography (GC) coupled with mass spectrometry (MS) or high resolution accurate mass (HRAM) mass spectrometry. LC-MS methods have been developed to cover a wider range of analytes which are not amenable by GC-MS methods. In particular, GC-MS methods cannot directly detect N-nitroso-N-methyl-4-aminobutyric acid (NMBA) and so sample derivatization is required, increasing sample preparation time and efforts. Moreover, LC-MS methods offer a suitable solution when ranitidine is tested for N-nitrosamine impurities. However, the FDA reported that possible degradation effects can occur when ranitidine is stored or analyzed at high temperatures resulting in subsequent formation of NDMA.<sup>6</sup>

Thermo Fisher Scientific supports pharmaceutical testing laboratories with comprehensive chromatographic solutions including LC-MS/MS, LC-HRAM, GC-MS, GC-MS/MS and high resolution accurate mass Orbitrap GC-MS, for targeted and untargeted analysis of nitrosamine impurities, in compliance with FDA guidelines. This guide will focus on GC-MS solutions and the LC-MS information can be found on the dedicated Nitrosamine resource page.



Analysis of nitrosamines using GC-MS coupled to headspace sampling

Thermo Scientific<sup>™</sup> ISQ<sup>™</sup> 7000 GC-MS system with Thermo Scientific<sup>™</sup> TriPlus<sup>™</sup> 500 Headspace Autosampler

#### Key features

- Headspace sampling with no sample preparation and low chemical background
- Headspace autosampler with up to 240 vial capability for increased productivity
- Thermo Scientific<sup>™</sup> Extractabrite<sup>™</sup> ion source with Thermo Scientific<sup>™</sup> NeverVent<sup>™</sup> technology for quick preventive maintenance without venting the MS ensuring maximum uptime
- Timed-SIM method set up for optimized selectivity and sensitivity

 Thermo Scientific<sup>™</sup> Chromeleon<sup>™</sup> Chromatography Data System (CDS) for instrument control, method development, quantitative analysis and reporting with a full suite of compliance-ready features to ensure data security, data integrity, and 21 CFR Part 11 compliance for operation in environments governed by Current Good Manufacturing Practice (cGMP) regulations.

For the determination of the most volatile nitrosamine impurities, headspace sampling technique is preferred as no sample preparation is required. One of the advantages of using this technique is that it removes the complexity of the matrix while improving the selectivity for the compounds of interest and reducing the risk of false positive results. Thermo Scientific TriPlus 500 HS autosampler coupled with Thermo Scientific<sup>™</sup> ISQ<sup>™</sup> 7000 single guadrupole GC-MS system achieves the required sensitivity with calculated LOD (S/N>3) of 0.004 ppm for NDMA and 0.009 ppm for NDEA and LOQ (S/N>10) =0.015 ppm for NDMA and 0.030 ppm for NDEA for the detection of nitrosamines at trace levels exceeding the FDA nitrosamine method requirements (LOD = 0.005 ppm for NDMA and 0.020 ppm for NDEA and LOD = 0.100 ppm for NDMA and 0.050 ppm for NDEA).<sup>5</sup>

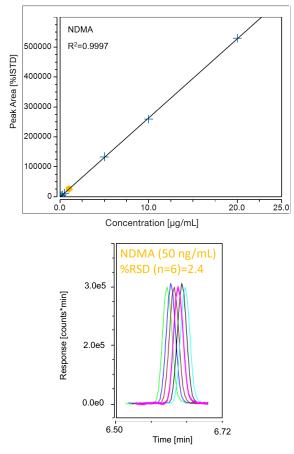


Figure 1. Examples of linearity for NDMA ranging from 0.03 to 20  $\mu g/mL$  and repeatability at 50 ng/mL in solvent standard

# Determination of nitrosamines using triple quadrupole GC-MS/MS coupled to direct liquid injection



Thermo Scientific<sup>™</sup> TSQ<sup>™</sup> 9000 GC-MS/MS system with Thermo Scientific<sup>™</sup> AS<sup>™</sup> 1310 Autosampler and Thermo Scientific TriPlus 500 Headspace Autosampler

#### Key features

- AS 1310 autosampler with up to 155 vials for increased sample throughput
- ExtractaBrite ion source with NeverVent technology for quick preventive maintenance without the need for venting the MS ensuring maximum uptime
- Auto-SRM and Time-SRM for fast method set up
- SRM selectivity reduces matrix interferences and enables low detection limits
- Chromeleon CDS for instrument control, method development, quantitative analysis and reporting in compliance-ready, enterprise, cGMP environments.

The published FDA methods include both liquid and gas chromatography coupled to mass spectrometry to ensure high selectivity and low limits of detection. Single or triple quadrupole mass analyzers provide acceptable selectivity to separate the analytes from the chemical background by the use of single ion monitoring (SIM) or selected reaction monitoring (SRM). Direct liquid injection coupled with triple quadrupole is the method of choice for determination of nitrosamines. Thermo Scientific<sup>™</sup> TSQ<sup>™</sup> 9000 triple quadrupole GC-MS/MS system coupled with Thermo Scientific<sup>™</sup> AS<sup>™</sup> 1310 Autosampler delivers high selectivity for detection and accurate quantitation of NDMA, NDEA, NEIPA, NDIPA and NDBA with linearity and sensitivity, exceeding the FDA requirements.<sup>5</sup>

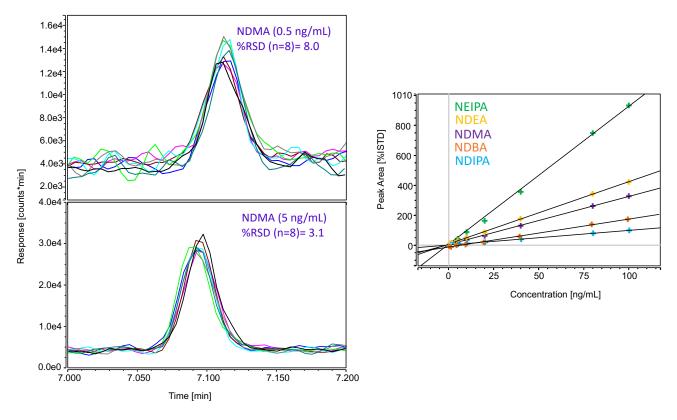


Figure 2. Examples of repeatability for NDMA at 0.5 ng/mL and 5.0 ng/mL in solvent standard and linearity for NDMA, NDEA, NEIPA, NDIPA and NDBA (1–100 ng/mL)

Table 1. Calculated limit of detection (LOD) and limit of quantification (LOQ), correlation coefficient (R<sup>2</sup>) and average calibration factors (AvCF %RSD) obtained from eight calibration levels (1–100 ng/mL)

| Nitrosoamine | Retention  | FDA LOD<br>Requiren | nent (ppm) | Calculated<br>_ LOD (S/N>3) | FDA LOQ<br>Requirement (ppm) |       | Calculated<br>LOQ (S/N>10) | Coefficient of                | AvCF |
|--------------|------------|---------------------|------------|-----------------------------|------------------------------|-------|----------------------------|-------------------------------|------|
| Impurity     | Time (min) | API                 | FPP        | (ppm)                       | ΑΡΙ                          | FPP   | (ppm)                      | Correlation (R <sup>2</sup> ) | %RSD |
| NDMA         | 7.11       | 0.005               | 0.008      | 0.0002                      | 0.008                        | 0.013 | 0.0005                     | 0.9997                        | 3.0  |
| NDEA         | 7.69       | 0.001               | 0.002      | 0.0002                      | 0.005                        | 0.008 | 0.0005                     | 0.9999                        | 2.0  |
| NEIPA        | 7.95       | 0.001               | 0.002      | 0.0005                      | 0.005                        | 0.008 | 0.001                      | 0.9997                        | 3.3  |
| NDIPA        | 8.14       | 0.001               | 0.002      | 0.0002                      | 0.005                        | 0.008 | 0.0005                     | 0.9998                        | 2.6  |
| NDBA         | 9.65       | 0.01                | 0.016      | 0.0005                      | 0.025                        | 0.04  | 0.01                       | 0.9988                        | 5.1  |

"Dealing with nitrosamines impurities in pharmaceutical industry is not a challenge if you have HS-GC-MS/MS TSQ technology. Selectivity and sensitivity to quantitate target volatile nitrosamines at low levels are the key to assure compliance now and for the future."

Anticipating future regulatory requirements with increased sensitivity from advanced electron ionization (AEI) ion source

#### Key features

 AEI ion source provides improved sensitivity with a more efficient ionization and improved robustness

As regulations are evolving quickly and limits become stringent, TriPlus 500 HS autosampler coupled with TSQ 9000 triple quadrupole GC-MS/MS system equipped with AEI technology provides the sensitivity performance to meet the current regulation and to future-proof against any reduction in limits of detection. The innovative design of the AEI ion source provides a highly efficient ionization of analytes and a more tightly focused ion beam, lowering the detection limits to < 0.5 ng/mL (S/N > 3).

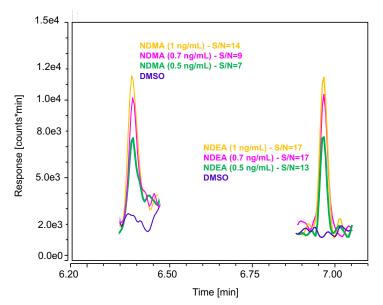


Figure 3. Peak-to-peak S/N for NDMA and NDEA spiked at 0.5, 0.7 and 1 ng/mL

- Dr. Siva Lakshmi (Laurus Labs, India)

## Improved analytical precision with the TriPlus 500 HS Autosampler

#### Key features

 Precise pneumatic control, short and inert sample path for highly reliable analyte transfer and reproducible results

When analyzing nitrosamines it is essential that results are consistent. If the analysis produces variable results it could cause uncertainty, which in turn could lead to the reanalysis of samples or even worse a false negative. The highly efficient pneumatic control and the short and inert sample path of the TriPlus 500 HS Autosampler ensure reliable and reproducible analyte injection and transfer, offering outstanding repeatability and precision in routine analysis at ultra-trace concentrations with peak area %RSD < 7 % at 0.7 ng/mL.

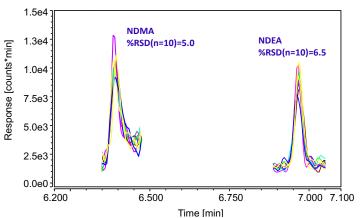


Figure 4. Overlaid injections (n=10) of NDMA (*m/z*=74>42) and NDEA (*m/z*=102>85) in solvent standard at 0.7 ng/mL (n=10)

"The Q Exactive GC mass spectrometer coupled with headspace sampler helps us a lot with confident identification of volatile organic compounds (VOCs) due to its excellent mass accuracy down to sub-1 ppm level."

– Dr. Dujuan Lu (SGS, USA)

#### Screening and quantification of genotoxic impurities in pharmaceuticals using Orbitrap GC-MS technology

The Thermo Scientific<sup>™</sup> Exactive<sup>™</sup> GC-MS system using Orbitrap<sup>™</sup> technology offers the advantage of full-scan (FS) operation with higher mass resolving power than single or triple quadrupoles and therefore, providing high levels of selectivity and quantitative performance. This enables not only target compounds to be confidently detected, but also widen the scope and identify additional compounds. Combined SIM and FS data acquisition at high resolution and high mass accuracy allow for targeted analysis of nitrosamine impurities with compliance to FDA method validation requirements,<sup>5</sup> as well as untargeted screening of contaminants with retrospective analysis capability.



Thermo Scientific<sup>™</sup> Exactive<sup>™</sup> GC Orbitrap<sup>™</sup> GC-MS system with Thermo Scientific<sup>™</sup> TriPlus<sup>™</sup> RSH Autosampler

#### **Orbitrap GC-MS coupled to direct liquid injection** Key features

- High resolving power and sub-1-ppm accurate mass for improved selectivity, lower limits of detection and confident compound identification
- Combined SIM and FS acquisition for targeted analysis of known impurities and untargeted screening for a broader and deeper understanding of samples
- Sample handling flexibility with TriPlus RSH autosampler offering liquid, syringe-based HS and dynamic headspace (ITEX-DHS) for further sensitivity
- ExtractaBrite ion source with NeverVent technology for quick preventive maintenance without venting the MS ensuring maximum uptime
- Chromeleon CDS for instrument control, method development, quantitative analysis and reporting in compliance-ready, enterprise, cGMP environments.

Orbitrap technology ensures high selectivity with very low background noise for the detection of ultra-trace level impurities with high confidence in compound identification. Impurities are identified based on the retention time, accurate mass information (±5 ppm mass error) and the characteristic fragment ion. Moreover, the elemental composition of the quantification ions can be used to check the isotopic pattern fit (measured versus theoretical). The Exactive GC-MS coupled with TriPlus RSH autosampler for liquid injection delivers excellent sensitivity with linearity ranging from 0.1 to 50 ng/mL and average

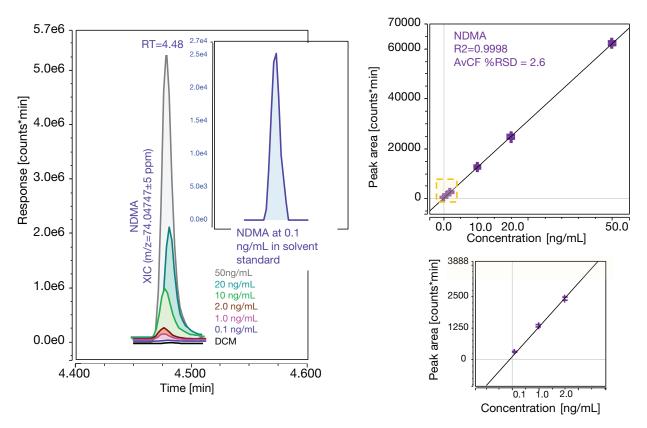


Figure 5. Linearity for NDMA (m/z=74.04747 ± 5 ppm) ranging from 0.1 to 50 ng/mL with R<sup>2</sup> = 0.9998 and AvCF %RSD=2.6



Thermo Scientific Q Exactive GC Orbitrap GC-MS/MS system with Thermo Scientific TriPlus 500 Headspace Autosampler

#### **Orbitrap GC-MS coupled to headspace sampling** Key features

- High resolving power and sub-1 ppm accurate mass for improved selectivity and confident compound identification
- Headspace sampling for easier sample preparation and lower chemical noise
- Headspace autosampler with up to 240 vial capability for increased productivity
- ExtractaBrite ion source with NeverVent technology for quick preventive maintenance without venting the MS ensuring maximum uptime
- Chromeleon CDS for instrument control, method development, quantitative analysis and reporting in compliance-ready, enterprise, cGMP environments.

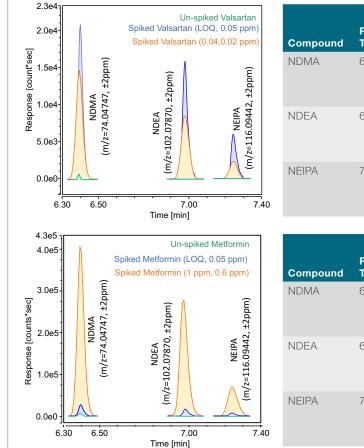
#### calibration factor %RSD=2.6.

The high selectivity and accurate mass (sub-1 ppm) of the Orbitrap technology add confidence in compound identification and ensure reliable quantitative performance irrespectively of the sample concentration. Quantitative analysis of un-spiked and spiked Valsartan and Metformin samples showed calculated amounts within ±15% of the spiked concentration with % recovery ranging from 80–120% and consistent sub-1-ppm mass accuracy.

## Untargeted screening of impurities in pharmaceutical substances

Untargeted screening and retrospective analysis can be carried out broadening the scope of the analysis. FS

data are acquired at 60,000 resolution (Full Width at Half Maxima measured at *m/z* 200) (1), deconvoluted with Thermo Scientific<sup>™</sup> TraceFinder<sup>™</sup> CDS and chromatographic peaks are putatively identified based on spectral library match against NIST 17 nominal mass library (2). The elemental composition and the mass accuracy information are used to confirm the molecular ion (3). Moreover, the isotopic patter match (measured vs theoretical) adds confidence in compound identification (4). Benzene and other residual solvent impurities could be detected in the Metformin sample spiked with some Class 1 and Class 2A residual solvents at 1/5 the concentration limits (Appendix 2) reported in the USP <467> method.<sup>7</sup>



|          | Retention  |        | ilsartan<br>ount (ppm) | _ Recovery | Mass Accuracy |  |
|----------|------------|--------|------------------------|------------|---------------|--|
| Compound | Time (min) | Spiked | Measured               | (%)        |               |  |
| NDMA     | 6.40       | -      | 0.0008                 | -          | -0.1          |  |
|          |            | 0.04   | 0.036                  | 90         | 0.6           |  |
|          |            | 0.05   | 0.044                  | 88         | 0.4           |  |
| NDEA     | 6.98       | -      | -                      | -          | -             |  |
|          |            | 0.04   | 0.034                  | 85         | 0.2           |  |
|          |            | 0.05   | 0.052                  | 104        | 0.7           |  |
| NEIPA    | 7.24       | -      | -                      | -          | -             |  |
|          |            | 0.02   | 0.022                  | 110        | 0.6           |  |
|          |            | 0.05   | 0.048                  | 96         | 0.4           |  |
|          |            |        |                        |            |               |  |

| Retention |            |        | etformin<br>unt (ppm) | _ Recovery |               |  |
|-----------|------------|--------|-----------------------|------------|---------------|--|
| Compound  | Time (min) | Spiked | Measured              | (%)        | Mass Accuracy |  |
| NDMA      | 6.40       | -      | 0.01                  | -          | 0.4           |  |
|           |            | 0.05   | 0.056                 | 112        | 0.7           |  |
|           |            | 1.0    | 1.05                  | 105        | 0.5           |  |
| NDEA      | 6.98       | -      | -                     | -          | -             |  |
|           |            | 0.05   | 0.056                 | 112        | 0.4           |  |
|           |            | 1.0    | 0.99                  | 99         | 0.1           |  |
| NEIPA     | 7.24       | -      | -                     | -          | -             |  |
|           |            | 0.05   | 0.054                 | 108        | 0.7           |  |
|           |            | 0.60   | 0.65                  | 109        | 0.3           |  |

Figure 6. Quantitative performance assessed for un-spiked and spiked Valsartan and Metformin samples with calculated concentrations within 15% the spiked amount, recoveries ranging from 80% to 120% and mass accuracy consistently < 1 ppm. Samples spiked at the FDA required LOQ (0.05 ppm), below the LOQ (0.04 and 0.02 ppm) and above the LOQ (1.0 and 0.6 ppm)

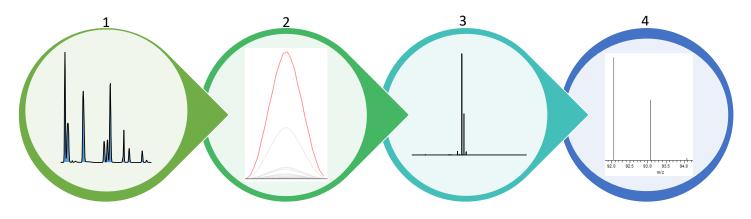


Figure 7. Workflow for unknown screening of volatile impurities. FS data are acquired at 60,000 resolution (FWHM at *m/z* 200) (1), chromatographic peaks are deconvoluted and putatively identified based on spectral library match against NIST 17 nominal mass library (2). The elemental composition and the mass accuracy information are used to confirm the molecular ion (3). Moreover, the isotopic pattern match (measured vs theoretical) adds confidence in compound identification (4)

| Compound                 | Retention<br>Time (min) | Reference <i>m/z</i>              | Theoretical <i>m/z</i> | Total<br>Score                          | Mass<br>Accuracy<br>(ppm)     |
|--------------------------|-------------------------|-----------------------------------|------------------------|---|-------------------------------|
| 1,1-Dichloroethene       | 2.76                    | 95.95286                          | 95.95281               | 98                                      | 0.50                          |
| Benzene                  | 3.39                    | 78.04639                          | 78.04640               | 99                                      | 0.13                          |
| cis-1,2-Dichloroethene   | 3.74                    | 95.95282                          | 95.95281               | 98                                      | 0.18                          |
| Toluene                  | 4.17                    | 92.06202                          | 92.06205               | 97                                      | -0.32                         |
| 1,4-Dioxane              | 4.32                    | 88.05189                          | 88.05188               | 95                                      | 0.11                          |
| Ethylbenzene             | 4.88                    | 106.0777                          | 106.07770              | 97                                      | 0.30                          |
| <i>p-</i> Xylene         | 4.94                    | 106.0776                          | 106.07770              | 97                                      | -0.44                         |
| <i>m-</i> Xylene         | 5.00                    | 106.0776                          | 106.07770              | 97                                      | -0.58                         |
| o-Xylene                 | 5.37                    | 106.0778                          | 106.07770              | 97                                      | 0.91                          |
| Chlorobenzene            | 5.62                    | 112.00752                         | 112.00743              | 96                                      | 0.79                          |
| B 2.50E+006              |                         |                                   | С                      | Measure                                 | d spectrum                    |
| 2.00E+006<br>1.50E+006   |                         |                                   | 100<br>80<br>60        | m,<br>C <sub>6</sub>                    | /z=78.04646<br>H <sub>6</sub> |
| 1.00E+006<br>5.00E+005   |                         |                                   | 40<br><u>9</u> 20      |   |                               |
| 0.00E+000<br>3.318 3.355 | 3.393 3.430             | 3.468 3.50<br>Theoretical Mass Er | -20                    | • |                               |

| 5.51              | 5.555               | 3.333 3.430      | 3.400                     | 5.565               | -20  |                  |    |
|-------------------|---------------------|------------------|---------------------------|---------------------|------|------------------|----|
| Component<br>Name | Acquired <i>m/z</i> | Fragment ID      | Theoretical<br><i>m/z</i> | Mass Error<br>(ppm) | -40  |                  |    |
| Benzene           | 78.04639            | C6H6             | 78.0464                   | 0.179381            |      |                  |    |
| Benzene           | 79.04982            | C(12)5 C(13)1 H6 | 79.04976                  | 0.812146            | -60  |                  |    |
| Benzene           | 78.04194            | C(12)5 C(13)1 H5 | 78.04193                  | 0.105072            |      |                  |    |
| Benzene           | 77.03854            | C6H5             | 77.03858                  | 0.415376            | -80  |                  |    |
| Benzene           | 77.03412            | C(12)5 C(13)1 H4 | 77.03411                  | 0.171353            | -100 |                  |    |
| Benzene           | 76.03072            | C6H4             | 76.03075                  | 0.35512             | 30   | 74               | 85 |
| Benzene           | 75.02304            | C6H3             | 75.02293                  | 1.532862            |      |                  |    |
| Benzene           | 74.01508            | C6H2             | 74.0151                   | 0.243194            |      | Library spectrum |    |
| Benzene           | 73.00724            | C6H              | 73.00728                  | 0.493102            |      |                  |    |

Figure 8. Example of deconvolution result table listing some of the compounds spiked in Metformin sample (A), deconvolution results with annotated fragments for peak eluting at RT=3.93 min and putatively identified as benzene based on NIST 17 library match, SI = 931 (B), spectrum comparison (measured vs library) (C)

#### Summary

The low levels at which the nitrosamine impurities could be present in pharmaceutical products pose challenges for analytical laboratories. The limits imposed by the current regulations are strict and could potentially be lowered in the future to ensure patients safety. Thermo Fisher Scientific offers a complete analytical solution for nitrosamine testing using LC-MS and GC-MS to cover a large range of pharmaceutical product types.

Highly versatile sample introduction configurations with liquid injection, static headspace (valve and loop and syringe based) and dynamic headspace (ITEX-DHS) coupled to single quadrupole, triple quadrupole and Orbitrap mass spectrometers are available to meet the current regulatory requirements and to address the future analytical needs.

Single quadrupole ISQ 7000 GC-MS and triple quadrupole TSQ 9000 GC-MS/MS systems are compliance-ready with FDA regulation, meeting or often exceeding the method requirements and providing the sensitivity, selectivity and precision needed for reliable quantitative analysis of these impurities.

It is critically important that the manufacturers of pharmaceutical products are assessing their products for any condition that might potentially lead to nitrosamine formation. Although quantification of known compounds, such as nitrosamines and precursors is essential, screening for unknown precursor chemicals that may contribute to the formation of nitrosamines is equally important. For this, Orbitrap GC-MS system combines the quantitative and qualitative strengths in one single platform allowing for targeted and untargeted screening of impurities and a deeper characterization of samples.

The high resolving power, consistent sub-1 ppm mass accuracy and the wide dynamic range of the GC-Orbitrap technology allows for fast and confident detection and identification of volatile impurities in pharmaceutical products regardless of concentration or complexity of the matrix. Compliance-ready Chromeleon CDS software offers fullyintegrated instrument control for streamlined workflows. This software delivers superior data integrity and security tools, networking capabilities, instrument control, automation, and data processing in compliance to the 21 Code of Federal Regulations (CFR) part 11.

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- International Agency for Research on Cancer; "IARC Monographs on the Evaluation of Carcinogenic Risks to Humans: Smokeless Tobacco and Some Tobacco-specific N-Nitrosamines, volume 89, 200
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Combined Direct Injection N-Nitrosodimethylamine (NDMA), N-Nitrosodiethylamine (NDEA), N-Nitrosoethylisopropylamine (NEIPA), N-Nitrosodiisopropylamine (NDIPA), and N-Nitrosodiibutylamine (NDBA), Impurity Assay by GC-MS/MS , 19/04/2019, https://www.fda.gov/media/123409/download

Combined Headspace N-Nitrosodimethylamine (NDMA), N-Nitrosodiethylamine (NDEA), N-Nitrosoethylisopropylamine (NEIPA), and N-Nitrosodiisopropylamine (NDIPA) Impurity Assay by GC-MS/MS, 29/04/2019, https://www.fda.gov/media/124025/download

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- Important Information about NDMA impurities in ranitidine products, https://www.fda.gov/drugs/drug-safety-and-availability/ questions-and-answers-ndma-impurities-ranitidine-commonly-known-zantac
- 7. USP <467> Organic Volatile impurities, Chemical Tests, United States Pharmacopeia, Interim Revision Announcement Official November 1, 2019; Official December 1, 2020

GC-MS, GC-MS/MS and GC-Orbitrap analytical methods

| TriPlus 500 HS Autosampler Parameters        | ;        |
|--|----------|
| Incubation Temperature (°C)                  | 150      |
| Incubation Time (min)                        | 15       |
| Vial Shaking                                 | Fast     |
| Vial Pressurization Mode                     | Pressure |
| Vial Pressure (kPa) (Auxiliary Gas Nitrogen) | 130      |
| Vial Pressure Equilibration Time (min)       | 1        |
| Loop Size (mL)                               | 1        |
| Loop/Sample Path Temperature (°C)            | 180      |
| Loop Filling Pressure (kPa)                  | 70       |
| Loop Equilibration Time (min)                | 1        |
| Needle Purge Flow Level                      | 2        |
| Injection Mode                               | Standard |
| Injection Time (min)                         | 1        |

| ISQ 7000 GC-MS Parameters      |               |
|--------------------------------|---------------|
| Transfer Line Temperature (°C) | 250           |
| Ionization Type                | El            |
| Ion Source                     | ExtractaBrite |
| Ion Source Temperature (°C)    | 300           |
| Electron Energy (eV)           | 70            |
| Aquisition Mode                | t-SIM         |

| ISQ 7000 GC-MS Parameters for t-SIM |                             |                        |  |  |  |  |
|-------------------------------------|-----------------------------|------------------------|--|--|--|--|
| Compound                            | <b>Retention Time (min)</b> | SIM Ion ( <i>m/z</i> ) |  |  |  |  |
| NDMA                                | 6.08                        | 74                     |  |  |  |  |
| NDMA                                | 6.08                        | 42                     |  |  |  |  |
| NDEA                                | 6.80                        | 102                    |  |  |  |  |
| NDEA                                | 6.80                        | 57                     |  |  |  |  |
| NDEA                                | 6.80                        | 42                     |  |  |  |  |



Method parameters: Thermo Scientific ISQ 7000 GC-MS system coupled with Thermo Scientific TriPlus 500 Headspace Autosampler

| Trace 1310 GC Parameters        |   |
|---------------------------------|---|
| Inlet Module and Mode           | SSL, split                                    |
| Split Ratio                     | 5:1   |
| Septum Purge Flow (mL/min)      | 5, constant                                   |
| Carrier Gas, Flow (mL/min)      | He, 1.0                                       |
| <b>Oven Temperature Program</b> |   |
| Temperature (°C)                | 45  |
| Hold Time (min)                 | 1   |
| Rate (°C/min)                   | 15  |
| Temperature 2 (°C)              | 180   |
| Rate (°C/min)                   | 20  |
| Temperature 3 (°C)              | 250   |
| Hold Time (min)                 | 1   |
| Column                          |   |
| Trace GOLD TG-WAXMS             | 30 m, 0.25 mm,<br>0.25 μm<br>(Ρ/Ν 26088-1420) |

Liquid Injection-GC-MS/MS

| AI/AS 1310 Autosampler Parameters |        |  |  |  |  |
|-----------------------------------|--------|--|--|--|--|
| Injection Volume (µL)             | 2      |  |  |  |  |
| Draw Speed                        | Slow   |  |  |  |  |
| Fill Stroke                       | 5      |  |  |  |  |
| Air Volume (µL)                   | 1      |  |  |  |  |
| Sample Depth                      | Bottom |  |  |  |  |
| Pre-Injection Delay Time (s)      | 2      |  |  |  |  |
| Post-Injection Delay Time (s)     | 3      |  |  |  |  |
| Pre-Injection Washing Cycles      | 5      |  |  |  |  |
| Post-Injection Washing Cycles     | 5      |  |  |  |  |
| Sample Wash Cycles                | 2      |  |  |  |  |

| TSQ 9000 GC-MS/MS Parameters   |               |
|--------------------------------|---------------|
| Transfer Line Temperature (°C) | 240           |
| Ionization Type                | El            |
| Ion Source                     | Extractabrite |
| Ion Source Temperature (°C)    | 230           |
| Electron Energy (eV)           | 70            |
| Aquisition Mode                | RM            |
| Detector Gain Multplier        | 3             |
| Emission Current (µA)          | 50            |

| Trace 1310 GC Parameters   |  |
|----------------------------|--|
|                            | 0.01                                       |
| Inlet Module and Mode      | SSL, splitless                             |
| Inlet Temperature (°C)     | 240  |
| Splitless Time (min)       | 1  |
| Split Ratio                | 25:1                                       |
| Septum Purge Flow (mL/min) | 5, constant                                |
| Carrier Gas, Flow (mL/min) | He, 1.0                                    |
| Oven Temperature Program   |  |
| Temperature (°C)           | 40   |
| Hold Time (min)            | 0.5  |
| Rate (°C/min)              | 20   |
| Temperature 2 (°C)         | 200  |
| Rate (°C/min)              | 60   |
| Temperature 3 (°C)         | 240  |
| Hold Time (min)            | 3  |
| Column                     |  |
| TR-WAX                     | 30 m, 0.25 mm,<br>1.0 μm<br>(P/N 260X296P) |

| TSQ 9000 GC-MS/MS Parameters for SRM |                                     |                                   |                |              |                             |
|--------------------------------------|-------------------------------------|-----------------------------------|----------------|--------------|-----------------------------|
| Compound                             | Precursor<br>Mass<br>( <i>m/z</i> ) | Product<br>Mass<br>( <i>m/z</i> ) | Start<br>(min) | End<br>(min) | Collision<br>Energy<br>(eV) |
| NDMA                                 | 74                                  | 42                                | 6.12           | 8.12         | 35                          |
| NDMA                                 | 74                                  | 44                                | 6.12           | 8.12         | 10                          |
| NDMA-d6                              | 80                                  | 50                                | 6.12           | 8.10         | 14                          |
| NDEA                                 | 102                                 | 44                                | 6.68           | 8.70         | 25                          |
| NDEA                                 | 102                                 | 56                                | 6.68           | 8.70         | 35                          |
| NDEA                                 | 102                                 | 85                                | 6.68           | 8.70         | 5                           |
| NEIPA                                | 116                                 | 70                                | 6.98           | 8.98         | 35                          |
| NEIPA                                | 116                                 | 99                                | 6.98           | 8.98         | 10                          |
| NDIPA                                | 130                                 | 42                                | 7.16           | 9.16         | 20                          |
| NDIPA                                | 130                                 | 88                                | 7.16           | 9.16         | 15                          |
| NDBA                                 | 158                                 | 99                                | 8.66           | 10.66        | 25                          |
| NDBA                                 | 158                                 | 116                               | 8.66           | 10.66        | 10                          |
| NUDA                                 | 100                                 | 110                               | 0.00           | 10.00        | 10                          |



Method parameters: Thermo Scientific TSQ 9000 GC-MS/MS system coupled with Thermo Scientific AS 1310 Autosampler

HS-GC-MS/MS (AEI)

#### TriPlus 500 HS Autosampler Parameters

| •  |          |
|--|----------|
| Incubation Temperature (°C)                  | 120      |
| Incubation Time (min)                        | 15       |
| Vial Shaking                                 | Fast     |
| Vial Pressurization Mode                     | Pressure |
| Vial Pressure (kPa) (Auxiliary Gas Nitrogen) | 115      |
| Vial Pressure Equilibration Time (min)       | 1        |
| Loop Size (mL)                               | 1        |
| Loop/Sample Path Temperature (°C)            | 180      |
| Loop Filling Pressure (kPa)                  | 52       |
| Loop Equilibration Time (min)                | 1        |
| Needle Purge Flow Level                      | 2        |
| Injection Mode                               | Standard |
| Injection Time (min)                         | 1        |
|  |          |

| TSQ 9000 GC-MS/MS Parameters   |     |
|--------------------------------|-----|
| Transfer Line Temperature (°C) | 220 |
| Ionization Type                | El  |
| Ion Source                     | AEI |
| Ion Source Temperature (°C)    | 250 |
| Electron Energy (eV)           | 70  |
| Aquisition Mode                | SRM |
| Detector Gain Multplier        | 10  |
| Emission Current (µA)          | 100 |
|                                |     |

| SSL, split                                   |
|--|
| 5:1  |
| 5, constant                                  |
| He, 1.0                                      |
|  |
| 40   |
| 0.5  |
| 20   |
| 160  |
| 10   |
| 220  |
| 1  |
|  |
| 30 m, 0.25 mm,<br>0.5 µm<br>(P/N 26086-2230) |
|  |

#### TSQ 9000 GC-MS/MS Parameters for SRM

| Compound | Precursor<br>Mass<br>( <i>m/z</i> ) | Product<br>Mass<br>( <i>m/z</i> ) | Start<br>(min) | End<br>(min) | Collision<br>Energy<br>(eV) |
|----------|-------------------------------------|-----------------------------------|----------------|--------------|-----------------------------|
| NDMA     | 74                                  | 42                                | 4.90           | 7.90         | 15                          |
| NDMA     | 74                                  | 44                                | 4.90           | 7.90         | 5                           |
| NDMA-d6  | 80                                  | 50                                | 4.88           | 7.88         | 5                           |
| NDMA-d6  | 80                                  | 46                                | 4.88           | 7.88         | 15                          |
| NDEA     | 102                                 | 44                                | 5.48           | 8.46         | 10                          |
| NDEA     | 102                                 | 85                                | 5.48           | 8.46         | 5                           |
| NEIPA    | 116                                 | 70                                | 5.74           | 8.74         | 35                          |
| NEIPA    | 116                                 | 99                                | 5.74           | 8.74         | 10                          |



Thermo Scientific TSQ 9000 GC-MS/MS system equipped with AEI ion source

Liquid Injection-Orbitrap-GC-MS

| TriPlus RSH Autosampler Parameters |     |
|------------------------------------|-----|
| Injection Volume (µL)              | 2   |
| Air Volume (µL)                    | 1   |
| Plunger Strokes                    | 7   |
| Filling Volume (µL)                | 3   |
| Pre-Injection Delay Time (s)       | 0   |
| Post-Injection Delay Time (s)      | 0   |
| Sample Pullup Speed (µL/s)         | 0.4 |
| Delay After Plunger Strokes (s)    | 1   |
| Viscosity Delay (s)                | 1   |
| Pre-Injection Washing Cycles       | 0   |
| Sample Rinse Volume(µL)            | 1   |
| Post-Injection Washing Cycles      | 4   |
| Washing Volume (µL)                | 3   |

| Exactive GC-MS Parameters for El         |                 |
|--|-----------------|
| Transfer Line Temperature (°C)           | 260             |
| Ionization Type                          | El              |
| Ion Source Temperature (°C)              | 230             |
| Electron Energy (eV)                     | 70              |
| Aquisition Mode                          | Full Scan       |
| Mass Range (Da)                          | 50-400          |
| Resolving Power (FWHM at <i>m/z</i> 200) | 60,000          |
| AGC Target                               | 1e <sup>6</sup> |
| Lockmass, Column Bleed                   | 207.03235       |

| Exactive GC-MS Parameters for SIM        |        |  |  |
|--|--------|--|--|
| Transfer Line Temperature (°C)           | 260    |  |  |
| Ionization Type                          | El     |  |  |
| Ion Source Temperature (°C)              | 230    |  |  |
| Electron Energy (eV)                     | 70     |  |  |
| Aquisition Mode                          | SIM    |  |  |
| Mass Range (Da)                          | 50-400 |  |  |
| Resolving Power (FWHM at <i>m/z</i> 200) | 30,000 |  |  |
| AGC Target                               | 5e4    |  |  |
| MSX Counts                               | 3      |  |  |
| Isolation Window ( <i>m/z</i> )          | 6      |  |  |

| - |               |                     |                |              |        |
|---|---------------|---------------------|----------------|--------------|--------|
| - | Exactive GC-I | MS Inclusion L      | ist for SI     | м            |        |
|   | Compound      | Mass ( <i>m/z</i> ) | Start<br>(min) | End<br>(min) | MSX ID |
| _ | NDMA-d6       | 80.08514            | 3.80           | 4.9          | 1      |
| - | NDMA          | 74.04747            | 3.80           | 4.9          | 2      |



Method parameters: Thermo Scientific Exactive GC-MS system coupled with Thermo Scientific TriPlus RSH Autosampler

| Trace 1310 GC Parameters   |  |
|----------------------------|--|
| Inlet Module and Mode      | SSL, splitless w/<br>Surge                     |
| Inlet Temperature (°C)     | 250  |
| Splitless Time (min)       | 1  |
| Split Flow (mL/min)        | 80   |
| Surge Pressure (kPa)       | 385  |
| Surge Duration (min)       | 1  |
| Septum Purge Flow (mL/min) | 5, constant                                    |
| Carrier Gas, Flow (mL/min) | He, 1.5  |
| Oven Temperature Program   |  |
| Temperature (°C)           | 35   |
| Hold Time (min)            | 3  |
| Rate (°C/min)              | 120  |
| Temperature 2 (°C)         | 270  |
| Hold Time (min)            | 1  |
| Column                     |  |
| TraceGOLD TG-1701MS        | 30.0 m, 0.25 mm,<br>0.5 μm<br>(P/N 26090-2230) |

HS-Orbitrap-GC-MS

| <b>TriPlus 500 HS Autosampler Parameters</b> |  |
|--|--|
| in ite eee ne nateeampier i arametere        |  |

| Incubation Temperature (°C)                  | 120      |
|--|----------|
| Incubation Time (min)                        | 15       |
| Vial Shaking                                 | Fast     |
| Vial Pressurization Mode                     | Pressure |
| Vial Pressure (kPa) (Auxiliary Gas Nitrogen) | 115      |
| Vial Pressure Equilibration Time (min)       | 1        |
| Loop Size (mL)                               | 1        |
| Loop/Sample Path Temperature (°C)            | 180      |
| Loop Filling Pressure (kPa)                  | 52       |
| Loop Equilibration Time (min)                | 1        |
| Needle Purge Flow Level                      | 2        |
| Injection Mode                               | Standard |
| Injection Time (min)                         | 1        |
|  |          |

| Trace 1310 GC Parameters   |  |  |
|----------------------------|--|--|
| Inlet Module and Mode      | SSL, split                                     |  |
| Split Ratio                | 5:1  |  |
| Septum Purge Flow (mL/min) | 5, constant                                    |  |
| Carrier Gas, Flow (mL/min) | He, 1.0  |  |
| Oven Temperature Program   |  |  |
| Temperature (°C)           | 40   |  |
| Hold Time (min)            | 0.5  |  |
| Rate (°C/min)              | 20   |  |
| Temperature 2 (°C)         | 160  |  |
| Rate (°C/min)              | 10   |  |
| Temperature 3 (°C)         | 220  |  |
| Hold Time (min)            | 1  |  |
| Column                     |  |  |
| Trace GOLD TG-WAXMS B      | 30.0 m, 0.25 mm,<br>0.5 μm<br>(P/N 26086-2230) |  |

| Exactive GC-MS Parameters for El         |                 |  |  |
|--|-----------------|--|--|
| Transfer Line Temperature (°C)           | 220             |  |  |
| Ionization Type                          | El              |  |  |
| Ion Source Temperature (°C)              | 250             |  |  |
| Electron Energy (eV)                     | 70              |  |  |
| Aquisition Mode                          | Full Scan       |  |  |
| Mass Range (Da)                          | 40-300          |  |  |
| Resolving Power (FWHM at <i>m/z</i> 200) | 60,000          |  |  |
| AGC Target                               | 1e <sup>6</sup> |  |  |
| Lockmass, Column Bleed                   | 207.03235       |  |  |

| Exactive GC-MS Parameters for SIM        |           |  |  |
|--|-----------|--|--|
| Transfer Line Temperature (°C)           | 220       |  |  |
| Ionization Type                          | El        |  |  |
| Ion Source Temperature (°C)              | 250       |  |  |
| Electron Energy (eV)                     | 70        |  |  |
| Aquisition Mode                          | Full Scan |  |  |
| Mass Range (Da)                          | 70–300    |  |  |
| Resolving Power (FWHM at <i>m/z</i> 200) | 30,000    |  |  |
| AGC Target                               | 5e4       |  |  |
| MSX Counts                               | 4         |  |  |
| Isolation Window ( <i>m/z</i> )          | 20        |  |  |

| Exactive GC-MS Inclusion List for SIM |                     |                |              |        |
|---------------------------------------|---------------------|----------------|--------------|--------|
| Compound                              | Mass ( <i>m/z</i> ) | Start<br>(min) | End<br>(min) | MSX ID |
| NDMA                                  | 74.04801            | 6.00           | 6.65         | 1      |
| NDEA                                  | 102.07931           | 6.70           | 7.20         | 2      |
| NEIPA                                 | 116.09496           | 7.00           | 7.40         | 3      |



Method parameters: Thermo Scientific Q Exactive GC-MS/MS system coupled with Thermo Scientific TriPlus 500 Headspace Autosampler

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#### Appendix 2

| Compound Name         | Concentration<br>Limit (ppm) |
|-----------------------|------------------------------|
| Class 1               |                              |
| 1,1-Dichloroethene    | 8                            |
| 1,1,1-Trichloroethane | 1500                         |
| Benzene               | 2                            |
| Carbon Tetrachloride  | 4                            |
| 1,2-Dichloroethane    | 5                            |

| Compound Name            | Concentration<br>Limit (ppm) |
|--------------------------|------------------------------|
| Class 2 A                |                              |
| Methanol                 | 3000                         |
| Acetonitrile             | 410                          |
| Dichloromethane          | 600                          |
| trans 1,2-Dichloroethene | 1870                         |
| cis 1,2-Dichloroethene   | 1870                         |
| Tetrahydrofuran          | 720                          |
| Cyclohexane              | 3880                         |
| Methycyclohexane         | 1180                         |
| 1,4-Dioxane              | 380                          |
| Toluene                  | 890                          |
| Chlorobenzene            | 360                          |
| Ehylbenzene              | 2170                         |
| <i>m</i> -Xylene         | 2170                         |
| p-Xylene                 | 2170                         |
| o-Xylene                 | 2170                         |

USP <467> concentration limits in ppm for Class 1 and Class 2A residual solvents

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