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Determination of Nitrosamine impurities in Losartan Potassium drug substance using Triple Quadrupole Liquid Chromatography Mass Spectrometry

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Introduction

The announcement for the recall of ARB medicines Valsartan, Losartan and Irbesartan made N-Nitroso impurities a focus for regulatory agencies including the FDA and the European Medicines Agency (EMA). Nitrosamine impurities are byproducts produced in trace amounts during the manufacturing processes of these medicines. These impurities/compounds are classified as probable carcinogens (i.e. potentially genotoxic impurities).

The liquid chromatography mass spectrometry-based method described in this poster was carried out on the 6470 Triple Quadrupole LC/MS (LC/TQ), presenting a comprehensive analysis of 6 nitrosamine impurities in Losartan Potassium drug substance at very low detection limits. All nitrosamine impurities are of very small molecular weight. These nitrosamine impurities include: N-nitrosodimethylamine (NDMA), N-nitrosodiethylamine (NDEA), N-nitroso-4-methyl-4-aminobutyric acid (NMBA), N-nitrosoethylisopropylamine (NEIPA), N-nitrosodiisopropylamine (NDIPA) and N-nitrosodibutylamine (NDBA).

Instrumentation

1290 Infinity II high-speed pump (G7120A)
 1290 Infinity II multisampler (G7167B)
 1290 Infinity II multicolumn thermostat (G7116B)
 1290 Infinity II variable wavelength detector (G7114B)
 6470 triple quadrupole LC/MS (G6470A)

Table 1: Instrumentation detail



Figure 1: 6470 triple quadrupole LC/MS

Experimental

Sample Preparation

The sample preparation procedure was optimized using the following steps.

1. Weigh 100mg(\pm 2mg) Losartan Potassium drug substance sample in a 15 mL centrifuge tube.
2. Add 5 mL sample diluent and vortex for 2minute.
3. Now put the sample in shaker at 450rpm for 40 minutes.
4. Centrifuge the sample at 5000 rpm for 10 minutes.
5. Filter the supernatant using 0.2 μ m nylon syringe filter into an LCMS vial.
6. Inject the sample into LC/TQ.

LC Conditions

| Needle wash | Methanol: Water/ 80:20 | | | | | | | | | | | | | | | | | | | | | | | | | | | |
|--------------------------|---|--------------|----|--------------|-----|---|------|-----|----|------|------|----|------|------|----|------|------|----|------|------|----|------|------|---|------|------|---|------|
| Sample diluent | Water: Methanol 95:5 | | | | | | | | | | | | | | | | | | | | | | | | | | | |
| Multisampler temperature | 6 °C | | | | | | | | | | | | | | | | | | | | | | | | | | | |
| Injection volume | 20 μ L | | | | | | | | | | | | | | | | | | | | | | | | | | | |
| Analytical column | Zorbax Eclipse Plus Phenyl-Hexyl, RRHD 2.1 x 100mm 1.8 μ m (P/N 959758-912) | | | | | | | | | | | | | | | | | | | | | | | | | | | |
| Column temperature | 40 °C | | | | | | | | | | | | | | | | | | | | | | | | | | | |
| Mobile phase A | 0.2 % formic acid in water | | | | | | | | | | | | | | | | | | | | | | | | | | | |
| Mobile phase B | Methanol | | | | | | | | | | | | | | | | | | | | | | | | | | | |
| Flow rate | 0.25 mL/min | | | | | | | | | | | | | | | | | | | | | | | | | | | |
| Gradient | <table border="1"> <thead> <tr> <th>Time (min)</th> <th>%B</th> <th>Flow(mL/min)</th> </tr> </thead> <tbody> <tr> <td>0.0</td> <td>5</td> <td>0.25</td> </tr> <tr> <td>5.0</td> <td>25</td> <td>0.25</td> </tr> <tr> <td>13.0</td> <td>55</td> <td>0.40</td> </tr> <tr> <td>20.0</td> <td>55</td> <td>0.40</td> </tr> <tr> <td>20.1</td> <td>95</td> <td>0.25</td> </tr> <tr> <td>23.0</td> <td>95</td> <td>0.25</td> </tr> <tr> <td>23.1</td> <td>5</td> <td>0.25</td> </tr> <tr> <td>25.0</td> <td>5</td> <td>0.25</td> </tr> </tbody> </table> | Time (min) | %B | Flow(mL/min) | 0.0 | 5 | 0.25 | 5.0 | 25 | 0.25 | 13.0 | 55 | 0.40 | 20.0 | 55 | 0.40 | 20.1 | 95 | 0.25 | 23.0 | 95 | 0.25 | 23.1 | 5 | 0.25 | 25.0 | 5 | 0.25 |
| Time (min) | %B | Flow(mL/min) | | | | | | | | | | | | | | | | | | | | | | | | | | |
| 0.0 | 5 | 0.25 | | | | | | | | | | | | | | | | | | | | | | | | | | |
| 5.0 | 25 | 0.25 | | | | | | | | | | | | | | | | | | | | | | | | | | |
| 13.0 | 55 | 0.40 | | | | | | | | | | | | | | | | | | | | | | | | | | |
| 20.0 | 55 | 0.40 | | | | | | | | | | | | | | | | | | | | | | | | | | |
| 20.1 | 95 | 0.25 | | | | | | | | | | | | | | | | | | | | | | | | | | |
| 23.0 | 95 | 0.25 | | | | | | | | | | | | | | | | | | | | | | | | | | |
| 23.1 | 5 | 0.25 | | | | | | | | | | | | | | | | | | | | | | | | | | |
| 25.0 | 5 | 0.25 | | | | | | | | | | | | | | | | | | | | | | | | | | |
| Stop time | 25 minutes | | | | | | | | | | | | | | | | | | | | | | | | | | | |
| Post time | 2 minutes | | | | | | | | | | | | | | | | | | | | | | | | | | | |

Table 2: LC conditions

Method Optimization

The 6470 LC/TQ was used for detecting the mass conditions for nitrosamine impurities in positive mode where M+H ion were found to be predominant precursor ions. The method was optimized using atmospheric pressure chemical ionization (APCI) source as most of the nitrosamines give better response and low noise background using APCI source. MRM method was converted into a dynamic MRM method.

MRM Transitions and Conditions

| Compound | Prec. Ion (m/z) | Product Ion (m/z) | Frag. (V) | CE (V) | CAV (V) | ± |
|----------|-----------------|-------------------|-----------|--------|---------|---|
| NDEA | 103.1 | 75.1 | 80 | 9 | 3 | + |
| NDEA | 103.1 | 47.1 | 80 | 17 | 3 | + |
| NDMA | 75.1 | 58 | 60 | 12 | 3 | + |
| NDMA | 75.1 | 43.1 | 60 | 18 | 3 | + |
| NMBA | 147.1 | 44.2 | 60 | 16 | 3 | + |
| NMBA | 147.1 | 87.2 | 60 | 10 | 3 | + |
| NEIPA | 117.1 | 75.1 | 75 | 8 | 3 | + |
| NEIPA | 117.1 | 47.1 | 75 | 18 | 8 | + |
| NDIPA | 131.1 | 89.1 | 75 | 6 | 3 | + |
| NDIPA | 131.1 | 43.1 | 75 | 12 | 8 | + |
| NDBA | 159.1 | 57.2 | 90 | 12 | 3 | + |
| NDBA | 159.1 | 41.1 | 90 | 22 | 3 | + |

Table 3: MRM transitions and conditions

| MS Conditions | |
|----------------------|---------|
| Gas Temperature | 300 °C |
| Gas Flow | 6 L/min |
| Capillary Voltage | 3000V |
| Nebulizer Pressure | 55 psi |
| APCI Heater | 350 °C |
| APCI Needle Positive | 4 µA |

Table 4: MS conditions

The chromatographic separation of Losartan Potassium drug substance and nitrosamine impurities was best achieved using Zorbax Eclipse Plus Phenyl-Hexyl column and diverter valve was programmed such that Losartan Potassium peak was diverted to waste and monitored using variable wavelength detector.

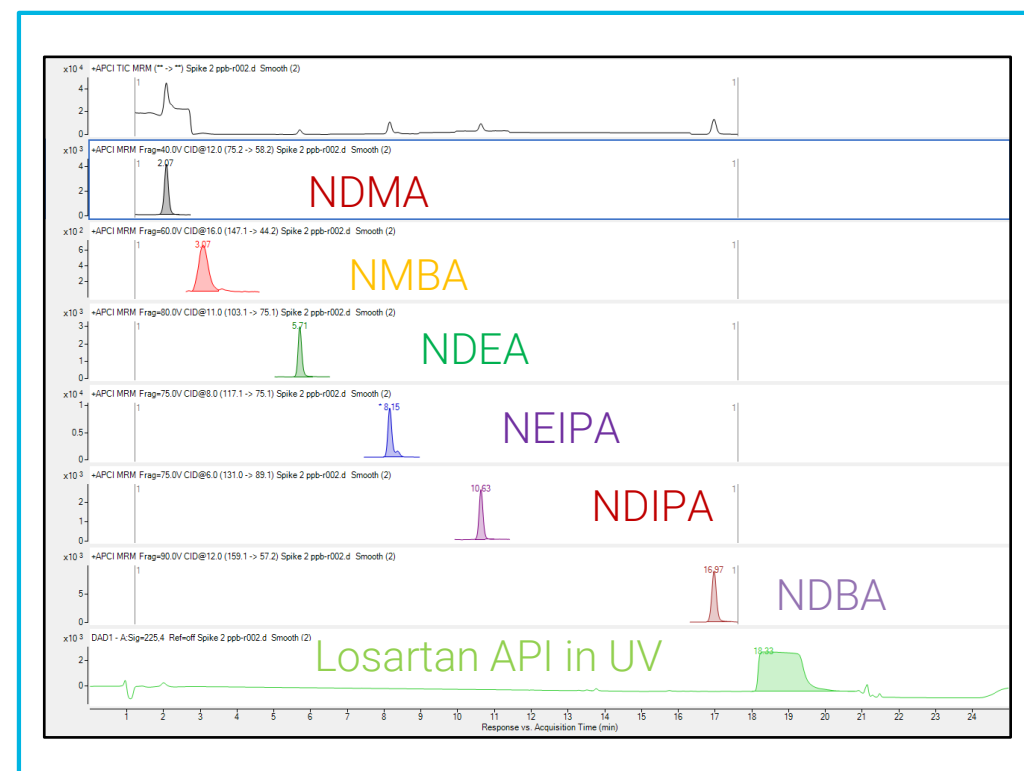


Figure 2: Representative EIC of NDMA, NMBA, NDEA, NEIPA, NDIPA and NDBA at 0.1 ppm conc. using 20mg/mL of Losartan Potassium API.

Below is presented the reproducibility data at 1ng/mL standard concentration for 8 replicates including bracketing standards (# 7 and 8) showing excellent peak area RSD % of < 6 % for each 6 nitrosamine impurities.

Area % RSD at 1ng/mL

| # | NDMA | NMBA | NDEA | NEIPA | NDIPA | NDBA |
|---------|-------|--------|--------|---------|-------|--------|
| 1 | 2556 | 5484 | 10530 | 36010 | 14023 | 18686 |
| 2 | 2409 | 5609 | 10727 | 36593 | 13478 | 18853 |
| 3 | 2436 | 4844 | 9962 | 34563 | 13899 | 16452 |
| 4 | 2442 | 4937 | 10067 | 32146 | 13871 | 16342 |
| 5 | 2435 | 4827 | 10066 | 32805 | 14375 | 16942 |
| 6 | 2578 | 4996 | 10182 | 32838 | 13822 | 16670 |
| 7 | 2442 | 4987 | 10145 | 33254 | 14335 | 16706 |
| 8 | 2434 | 4966 | 10193 | 33108 | 13868 | 16691 |
| Avg | 2467 | 5081 | 10234 | 33915 | 13959 | 17168 |
| SD | 63.16 | 295.66 | 259.96 | 1629.64 | 289.9 | 1005.5 |
| RSD (%) | 2.56 | 5.82 | 2.54 | 4.81 | 2.08 | 5.86 |

Table 5: Peak area % RSD for 8 replicates at 1ng/mL

Method Performance Characterization

Figure 3 shows the calibration curves for the standard calibration of all 6 nitrosamines. The relevant calibration range for NDMA, NMBA and NDEA is from 0.05ng/mL to 25ng/mL and for NEIPA, NDIPA and NDBA is from 0.1 ng/mL to 25ng/mL.

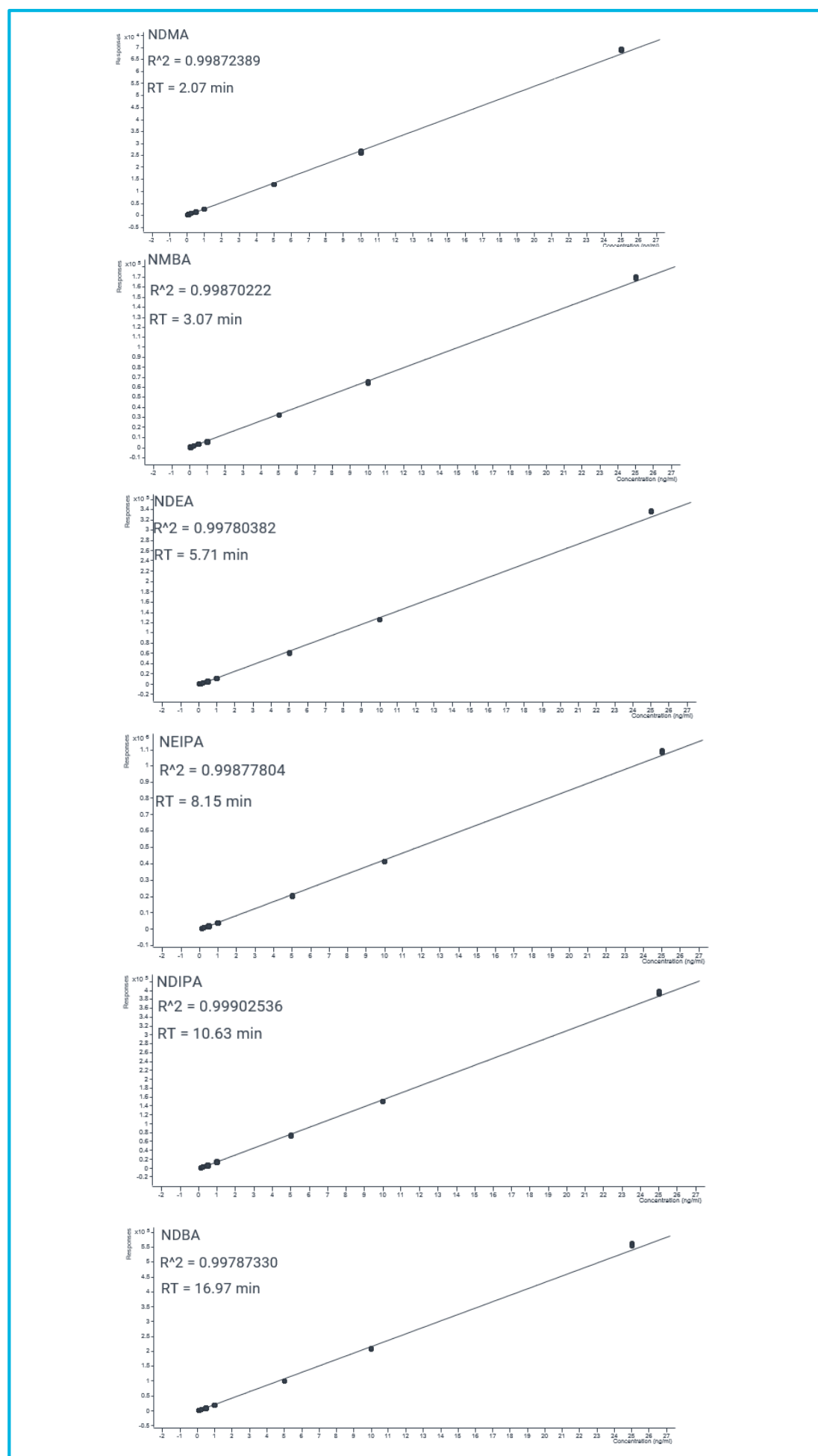


Figure 3: Calibration curves of all 6 nitrosamines with $r^2 > 0.997$

Recovery Study

The recovery experiment shows excellent recovery of $\pm 20\%$ of the spiked concentrations.

| Nitrosamine Impurity | Concentration (ng/mL) | Recovery % |
|----------------------|-----------------------|------------|
| NDMA | 2 | 110 |
| NMBA | 1 | 113 |
| NDEA | 1 | 103 |
| NEIPA | 1 | 100 |
| NDIPA | 1 | 98 |
| NDBA | 2 | 91 |

Table 6: Recovery data in Losartan API

Conclusions

- The method provides excellent reproducibility at USFDA defined LOQ concentrations levels as it shows area RSDs of $< 6\%$ with bracketing standards included in the calculations.
- The method is a ready to use method for analysis of Losartan Potassium drug substance batches as the method shows excellent recovery.
- The Losartan Potassium drug substance peak is chromatographically well separated from nitrosamine peaks so it can easily be diverted from the MS. Therefore, there is no contamination to the mass spectrometer due to a high concentration of API.

References