Poster Reprint

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# Analysis of Dioxins Utilizing Time-ofFlight for Low Level Quantitation 

Jeff Hollis, Matthew Curtis, Courtney Milner
Agilent Technologies, Santa Clara, CA USA

## Introduction

The World Health Organization categorizes dioxins as part of the "dirty dozen" of persistent organic pollutants. There are 210 congeners of dioxins, of these, the 17 with chlorine atoms in the $2,3,7,8$ positions are classified as highly toxic and require trace level detection. In addition to the toxicity, they bioaccumulate in the fat tissue of animals, with a halflife of 7-11 years. The food chain is the primary source of the main human exposure. In 2008, several tons of pork in Ireland was recalled and destroyed due to a dioxin contamination 200x the safe limit. The current GC/MS method designated to the analysis of the environmental dioxin contamination is EPA 1613B ${ }^{1}$, which requires specific chromatographic separation and high resolving power mass spectrometry for confident identifications.
The data presented in this poster illustrates the analytical capability of an accurate mass high resolution GC/Q-TOF for the analysis of dioxins. EPA method 1613b${ }^{1}$ has several data requirements to pass the criteria for a successful analysis of dioxins. The qualifying ion ratio cannot exceed $15 \%$ of theoretical, the resolving power must be >10,000 using the 10\% valley calculation, the mass accuracy must be within 5 ppm , and the lowest calibration standard (CS2) must have a $\mathrm{S} / \mathrm{N}$ value $>10$. The $7250 \mathrm{GC} / \mathrm{Q}-\mathrm{TOF}$ meets or exceeds each of these requirements.


Figure 1: Agilent 7250 GC/Q-TOF

## Experimental

## Sample Preparation:

Chemical standards were provided by Accustandard for the 5 -point calibration curve. 1613b prepped samples were provided by Eurofins TestAmerica (West Sacramento, CA) to evaluate the method with real-world extractions. Calibration curve and continuing calibration standards provided quantitative results for the extracted samples. Toluene and hexane were used for the autosampler solvent washes to reduce the amount of carry-over. Lockmass introduction is discussed below to meet the method criteria, but not necessary with this instrument because reduced mass assignment drift. Mass calibration between injections is a more productive approach for mass accuracy confidence. Analytical conditions for the GC/Q-TOF platform are listed in Table 1

## Lockmass Introduction:

The Agilent Thermal Separation Probe installed into the multimode inlet(MMI) delivered a consistent amount of perfluorokerosene(PFK) into the ion source throughout the analysis (Figure 2). The MMI was kept at a constant temperature of $50^{\circ} \mathrm{C}$, using clean, dry air and connected to the MS with a $10 \mathrm{~m} \times 100 \mu \mathrm{~m}$ deactivated tube which provided a flow of $0.05 \mathrm{~mL} \mathrm{~min}^{-1}$.

## Software:

All of the data analysis was performed with the MassHunter Suite. This included MassHunter Qualitative Analysis 10, and MassHunter Quantitative Analysis 10 using the SureMass peak detection algorithm.
Table 1: Agilent 7250 GC/Q-TOF; 7890B GC Parameters

| Column | DB-5ms, 60 m, 0.25 mm ID, 0.25 $\mu \mathrm{m}$ film |
| :---: | :---: |
| Injection volume and liner | $\begin{array}{ll}1 \mu \mathrm{~L} & 2 \mathrm{~mm} \text { straight liner; } \\ & \mathrm{UI}\end{array}$ |
| Pulsed Splitess | 40psi for 1 min; 0.9min purge |
| Inlet temperature | $290{ }^{\circ} \mathrm{C}$ |
| Oven temperature | $180^{\circ} \mathrm{C}$ for 1 min |
| program | $30^{\circ} \mathrm{C} / \mathrm{min}$ to $270^{\circ} \mathrm{C}$; hold 1 min |
|  | $2^{\circ} \mathrm{C} / \mathrm{min}$ to $310^{\circ} \mathrm{C}$ |
|  | $10^{\circ} \mathrm{C} / \mathrm{min}$ to $320^{\circ} \mathrm{C}$; hold 6 min |
| Carrier gas | Helium; $1.0 \mathrm{~mL} \mathrm{~min}{ }^{-1}$ |
| Transfer line temperature | $300{ }^{\circ} \mathrm{C}$ |
| Source temperature | $280^{\circ} \mathrm{C}$ |
| Quadrupole temperature | $150^{\circ} \mathrm{C}$ |
| Spectral range | 225 to $650 \mathrm{~m} / \mathrm{z}$ |
| Spectral acquisition rate | 3 Hz , both centroid and profile |
| Electron Energy | 70 eV |
| Emission | $6 \mu \mathrm{~A}$ |

## Results and Discussion



Figure 2: RTIC of CS3 with extracted Ion Chromatogram for two of the lockmass ions of PFK used for mass calibration.


Figure 3: FWHM resolving power observed (bottom number) for the CS3, mid-point calibration standard for 2,3,7,8-TCDD and 2,3,7,8-TCDF.


Figure 4: FWHM resolving power observed (bottom number) for the CS3, mid-point calibration standard for 1,2,3,4,6,7,8HpCDF and 1,2,3,4,6,7,8-HpCDD.


Figure 5: FWHM resolving power observed (bottom number) for the CS3, mid-point calibration standard for OCDF and OCDD.


Figure 6: 2,3,7,8-TCDD lowest calibration quantitation with S/N, mass accuracy and ISTD response


Figure 7: 2,3,7,8-TCDD calculated concentrations for several real-world extracted samples. CCs were ran 12-hours apart and provided calibration curve accuracy confidence.

Table 2: 20x dilution of CS5 was injected 8 times resulting in the following concentrations; $1 \mathrm{ng} \mathrm{mL}^{-1}$ TCDD \& TCDF, $2.5 \mathrm{ng} \mathrm{mL}^{-1}$ Penta through Hepta, $10 \mathrm{ng} \mathrm{mL}^{-1}$ OCDD \& OCDF. IDL was calculated using Student's t-test with a 99\% confidence.

|  |  |  |  |  |  |  |  |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Name | Area | Area | ea | Area | Area | Area | Area | Area | Average | STDev | RSD |  |
| 1,2,3,4,6,7,8-H | 1274 | 14565 | 1395 | 12710 | 11897 | 1308 | 12880 | 1367 | 13189.0 | 838.7 | 0.06 |  |
| 1,2,3,4 | 16479 | 22594 | 24337 | 23295 | 18331 | 24066 | 23362 | 530 | 22124.3 |  |  |  |
| 1,2,3,4,7,8,9-HpCD | 126 | 1816 | 17 | 15538 | 13 | 34 | 01 | 030 | 15541.0 |  |  |  |
| 1,2,3,4,7,8-HxCDD | 19719 | 23996 | 21407 | 21367 | 17502 | 19018 | 558 | 94 | 20314. |  |  |  |
| 1,2,3,4,7,8-HxCD | 298 | 36075 | 35077 | 32388 | 26850 | 31292 | 32669 | 32081 | 32038 | 2883 | 0 | 0.7 |
| 1,2,3,6,7,8-HxCDD | 19020 | 24943 | 20586 | 20511 | 17693 | 22928 | 22717 | 20818 | 21152 | 2309 |  | 0.8 |
| 1,2,3,6,7,8-HxCDF | 3261 | 36963 | 33059 | 33171 | 30183 | 32834 | 34837 | 34134 | 3347 |  |  | 0.4 |
| 1,2,3,7,8,9-HxCDD | 1843 | 22717 | 21028 | 19945 | 18239 | 19798 | 20398 | 19801 | 20045.4 |  |  | 0.5 |
| 1,2,3,7,8,9-HxCDF | 19897 | 24228 | 24604 | 22751 | 18558 | 21059 | 21556 | 24186 | 22 | 2215.3 | 30.09 |  |
| 1,2,3,7,8-PeCDD | 251 | 29071 | 27178 | 26650 | 23121 | 25881 | 26830 | 26840 | 26336 | 1724.0 |  |  |
| 1,2,3,7,8-PeCDF | 381 | 46071 | 42357 | 41075 | 36847 | 41165 | 43002 | 43448 | 41513.4 | 2949.5 | 5.07 | 0.5 |
| 3,4,6,7,8-HxCDF | 287 | 34708 | 32366 | 29751 | 25829 | 29509 | 30520 | 32064 | 30 | 2674 | 0.08 | 0.6 |
| 2,3,4,7,8-PeCDF | 40203 | 44270 | 39925 | 41165 | 35841 | 39991 | 38045 | 43575 | 40376 | 2736 | 0.06 | 0.5 |
| 2,3,7,8-TCDD | 6038 | 7842 | 8042 | 6614 | 5666 | 6767 | 5629 | 6705 | 6662.9 | 07. | 0.14 | 0.4 |
| 2,3,7,8-TCDF | 6685 | 8292 | 6805 | 6973 | 6715 | 6983 | 7115 | 7806 | 7171.8 | 575.2 | 0.07 | 0.2 |
| OCDD | 14844 | 18766 | 18196 | 15281 | 12669 | 17232 | 17223 | 17485 | 16462. | 2033.1 | 0.12 | 3.5 |
| OCDF | 16030 | 2206 | 20710 | 17 | 15176 | 19232 | 19661 | 21236 | 18909.4 | 2523.4 |  | 3.6 |

Table 3: Signal to noise for each analyte with respect to the calibration concentration. The lowest calibration standard is highlighted in blue to show each analyte is $>10$.

| Compound Method | CS2 |  | CS3 |  | CS4 |  | CS5 |  | CS6 |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Name | S/N | Resp. | S/N | Resp. | S/N | Resp. | S/N | Resp. | N | Resp. |
| [13C]1,2,3,4,6,7,8-HpCDD | 644.54 | 303229 | 264.75 | 260546 | 315.22 | 183432 | 346.74 | 205249 | 592.8 | 219234 |
| [13C]1,2,3,4,6,7,8, +HPCDF | 535.06 | 214919 | 689.23 | 190678 | 251.15 | 137157 | 299.78 | 159165 | 417 | 157652 |
| [13C]1,2,34,7,8.-HpCDF | 161.44 | 163451 | 362.06 | 138510 | 251.66 | 93991 | 247.95 | 104984 | 271.98 | 107817 |
| [13C] $1,2,3,4,7,8$ - HXCDD | 557.77 | 405837 | 485.88 | 363188 | 487.34 | 269649 | 429.28 | 300079 | 311.46 | 303614 |
| \|13Cl1,2,3,4,8-HxCDF | 650.44 | 371447 | 508.24 | 341398 | 551.27 | 257123 | 556.9 | 280229 | 680 | 274086 |
| [13C) $1,2,3,6,7,8$ - HXCDD | 490.39 | 405837 | 395.18 | 363188 | 370.91 | 269649 | 429.28 | 300079 | 386.35 | 303614 |
| [13Cl1,2,3,6,7,-HxCDF | 650.44 | 371447 | 429.38 | 341398 | 551.27 | 257123 | 556.9 | 280229 | 627.1 | 274086 |
| \|13C) $1,2,3,7,8,9-\mathrm{HxCDD}$ | 544.18 | 449604 | 447.78 | 407700 | 48.32 | 279405 | 448.23 | 317813 | 40.06 | 324780 |
| ${ }^{1} 13 \mathrm{Cl} 11,2,3,8,89-\mathrm{HXCDF}$ | 412.85 | 252947 | 428.05 | 239139 | 294.34 | 158896 | 355.44 | 177271 | 181.21 | 180705 |
| [13C]1,2,3,7,8-PeCDD | 1103.74 | 592545 | 1117.96 | 543791 | 732.06 | 392381 | 900.8 | 441348 | 1204.9 | 441402 |
| ${ }^{13} 13 \mathrm{C} 11,23,7,8$-PeCDF | 1597.76 | 939554 | 1490.02 | 873399 | 1075.49 | 655344 | 409.03 | 708179 | 1367.44 | 705442 |
| ${ }^{1} 13 \mathrm{C} \mid 2,3,4,6,7,8$ - $\mathrm{H} \times \mathrm{CDF}$ | 625.31 | 345507 | 613.38 | 318097 | 431.46 | 228667 | 603.55 | 252140 | 617.17 | 253215 |
| \|13C12,3,4,7.-PeCDF | 1539.43 | 940325 | 1083.29 | 847683 | 977.79 | 613773 | 1195.89 | 679789 | 1056.12 | 690386 |
| [13C]OCDD | 531.43 | 357078 | 423.91 | 280076 | 362.39 | 177791 | 438.15 | 206948 | 526.33 | 218232 |
| [13C]121,23,4-TCDD | 1037.59 | 569601 | 626.58 | 516957 | 905.26 | 429298 | 770.52 | 456612 | 786.03 | 447944 |
| ${ }^{\text {[13C] } 122,3,3,8-T C D D ~}$ | 982.06 | 538853 | 597.54 | 494540 | 814.1 | 381822 | 663.81 | 409987 | 860.71 | 401762 |
| ${ }^{13 C 1122,3,7,-T C D F}$ | 1761.94 | 838685 | 1014.73 | 786886 | 1086.92 | 573289 | 1246.15 | 635143 | 1303.45 | 626658 |
| [37C1142,3,7,8-TCDD | 16.12 | 6128 | 74.31 | 20761 | 319.95 | 88690 | 723.98 | 417377 | 3684.07 | 2424298 |
| 1,2,3,4,6,7,8-HpCDD | 30.38 | 8342 | 113.1 | 25388 | 190.05 | 89154 | 508.57 | 448096 | 3162.48 | 2743066 |
| 1,2,3,4,6,7,8-HpCDF | 49.82 | 11917 | 86.37 | 42207 | 288.07 | 160362 | 1140.15 | 863318 | 1009.44 | 4999904 |
| 1,2,3,4,7,8,9-HpCDF | 18.06 | 7752 | 75.59 | 30986 | 204.97 | 102956 | 522.91 | 512227 | 2702.56 | 3216108 |
| 1,2,3,4,7,8-HYCDD | 41.37 | 9544 | 72.82 | 36680 | 102.93 | 136063 | 635.01 | 711841 | 2305.15 | 4165972 |
| 1,2,3,4,7.8.- ${ }^{\text {CCDF }}$ | 32.39 | 16622 | 135.28 | 59161 | 477.9 | 237228 | 1336.66 | 1251005 | 5078.75 | 7396482 |
| 1,2,3,6,7,8-HYCDD | 41.37 | 9544 | 72.82 | 36680 | 230.63 | 136063 | 755.42 | 711841 | 2305.15 | 4165972 |
| 1,2,3,6,7,8HXCDF | 32.39 | 16622 | 135.28 | 59161 | 477.9 | 237228 | 1390.2 | 1251005 | 5078.75 | 7396482 |
| 1,2,3,7,8,9-HxCDD | 12.81 | 12237 | 72.85 | 36292 | 245.53 | 136074 | 681.62 | 696725 | 2306.57 | 4168375 |
| 1,2,3,7,8,-HXCDF | 47.41 | 10570 | 122.89 | 43991 | 307.94 | 152201 | 978.04 | 788726 | 2875.38 | 4720151 |
| 1,2,3,7,8-PeCDD | 35.98 | 12852 | 127.07 | 46936 | 427.96 | 186588 | 1339.36 | 969781 | 6920.23 | 5471838 |
| 1,23,7,8-PeCDF | 74.9 | 20449 | 253.63 | 74792 | 472.11 | 302353 | 609.53 | 1517045 | 6443.85 | 9012186 |
| 2,3,4,6,7,8-HCDF | 84.87 | 14033 | 163.71 | 56051 | 390.07 | 210199 | 1054.57 | 1138956 | 4484.87 | 6711514 |
| 23,4,7,8-PeCDF | 111.14 | 20032 | 223.94 | 70797 | 481.46 | 286077 | 1909.46 | 1473084 | 6437.96 | 8759891 |
| 2,3,7,8-TCDD | 19.03 | 2927 | 83.43 | 11761 | 119.69 | 45676 | 1270.22 | 214743 | 1692.75 | 1227079 |
| 23,7,8-TCDF | 20.23 | 3944 | 49.28 | 12398 | 170.45 | 52924 | 492.41 | 239347 | 347.06 | 1369294 |
| OCDD | 40.6 | 9557 | 26.1 | 25392 | 188.95 | 94465 | 732.05 | 474488 | 4756.47 | 3071322 |
| OCDF | 53.83 | 11633 | 79.6 | 34519 | 294.4 | 111974 | 789.57 | 592775 | 2937.81 | 3762547 |

For the qualifying ratios, an outlier is configured in MassHunter Quantitative analysis for each compound to flag if the sample has a deviation greater than $15 \%$, the largest difference was 10\% for the 10x dilution of the CS2 lowest calibration standard. The resolving power requirement of 10,000 by $10 \%$ valley converts to a requirement of $>20,000$ for FWHM (Figure 3). The data collects illustrated that the instrument was able to provide the required resolving power with the lowest calculation of $\sim 24,000$ and most were $>32,000$. The largest mass accuracy deviation observed was 2.4 ppm for the lowest calibration level while most were 1ppm or below. Eight replicate injections were performed with a diluted standard with varied concentrations of the different species. For the Tetra compounds the concentration was $1 \mathrm{ng} \mathrm{mL}^{-1}$, the penta- through heptawas $2.5 \mathrm{ng} \mathrm{mL}^{-1}$ and the octas were $10 \mathrm{ng} \mathrm{mL}^{-1}$ provided an instrument detection limit of 0.2-0.4, 0.4-0.9 and 3.53.6, respectively (Table 2). For the $\mathrm{S} / \mathrm{N}$ requirement of the lowest calibration standard, the lowest value was 12, for one compound and the largest was 111 and an average of 44 (Table 3). Contaminated samples were also analyzed to calculate the amount of 2,3,7,8-isomer dioxins were present (Figure 7).

## Conclusions

High resolving power, accurate mass and high sensitivity provided data to meet the performance criteria for EPA method 1613b.

- Time-of-flight spectral acquisition speeds provided accurate isotope intensities
- SureMass peak detection produced consistent mass accuracy
- Resolving power for the analytes exceeded the requirement of the method
- High resolving power quantitation provided excellent results for complex samples.


## Reference

${ }^{1}$ Telliard, W.A., 1994. Method 1613b Tetra- through OctaChlorinated Dioxins and Furans by Isotope Dilution HRGC/HRMS. U.S. Environmental Protection Agency, Office of Water.

